



**INRA**  
SCIENCE & IMPACT

# Molecular genetics of the adrenocortical axis and breeding for robustness

Pierre MORMEDE  
Elena E. TERENINA

National Institute for Agronomical Research  
Laboratory for Livestock Genetics, Physiology and Breeding (GenPhySE),  
Castanet-Tolosan, France



Diet, agriculture, environment

[pierre.mormede@inra.fr](mailto:pierre.mormede@inra.fr)  
[elena.mormede@inra.fr](mailto:elena.mormede@inra.fr)  
[www.mormede.com](http://www.mormede.com)

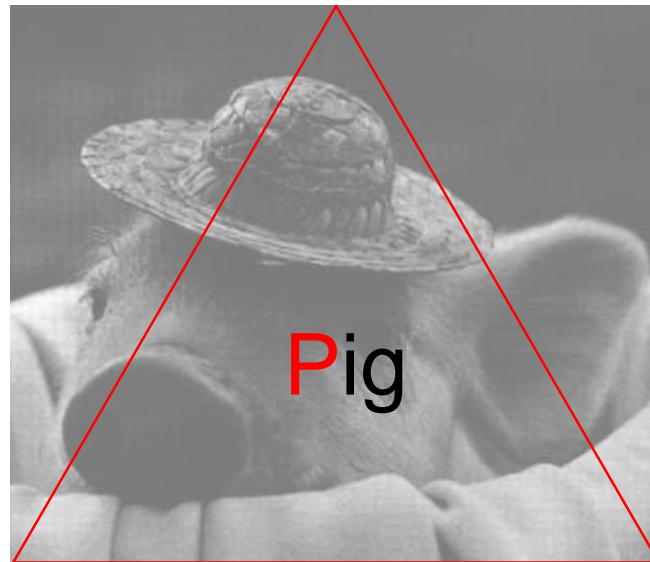
# New challenges for farm animal breeding

## Sustainability

- environmental footprint (GHG, N, P)
- antibiotics, heavy metals & hormones

## Profit

- Reduced production costs
  - feed efficiency
  - less labour (independent animals)
  - adaptability to environmental constraints



## Planet

- food safety
- product quality
- animal welfare
- rural lifestyle

## People

Main breeding objectives: feed efficiency, robustness, product quality

# ROBUSTNESS

- ❖ The concept of robustness refers to the ability of the animals to express their production potential in a wide range of environmental conditions without compromising their physical health and welfare (Knap 2005)
- ❖ Includes several components
  - Sensitivity of the production potential
    - ✓ to diverse commercial environments
    - ✓ and to external stressors (temperature)
  - Functional traits
    - ✓ quality of supportive tissues (e.g. leg soundness)
    - ✓ newborn survival
    - ✓ resistance to diseases (efficiency of the immune system )
    - ✓ productive longevity

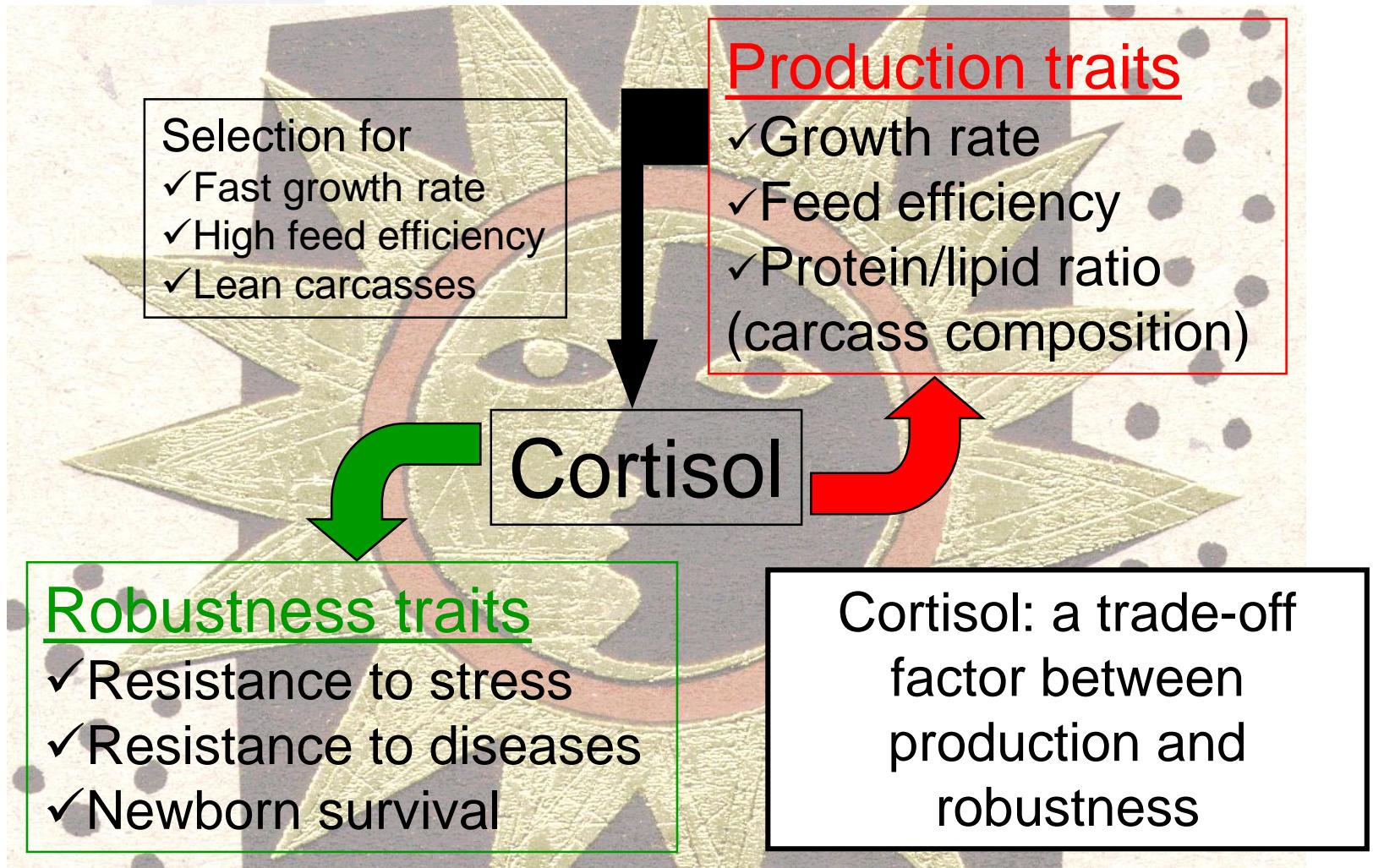
# Robustness as a breeding objective

‘Sustainable breeding goals combine robustness traits with production traits to such an extent that selection balances genetic change in production potential with genetic change in environmental sensitivity’ (Knap 2009)

## Selection strategies

1. Inclusion of directly measurable **functional traits** in the breeding objective and in the selection index (BLUP) – leg soundness, newborn survival rate, fertility, cellularity of milk ...
2. **Reaction norm** analysis – global sensitivity to the environment (Lynch and Walsh 1998) – low heritability
3. **Canalizing selection** for a reduced variance
4. Genetics of **stress responses**  
**behavioural and neuroendocrine**

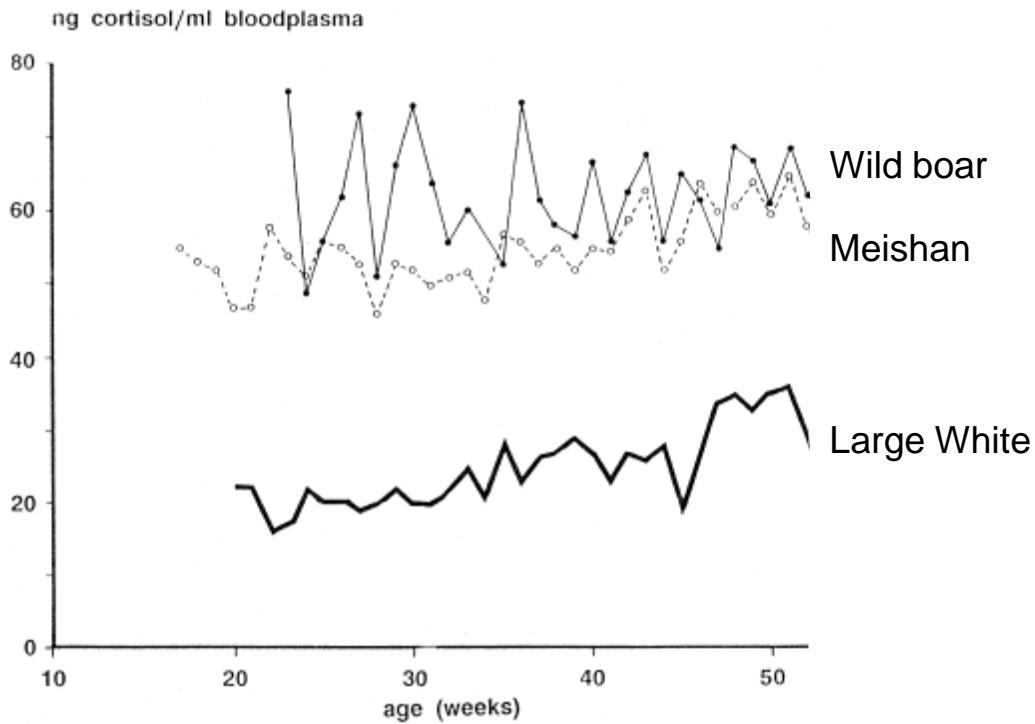
# Effects of cortisol on production and robustness traits





# Domestication and the HPA axis

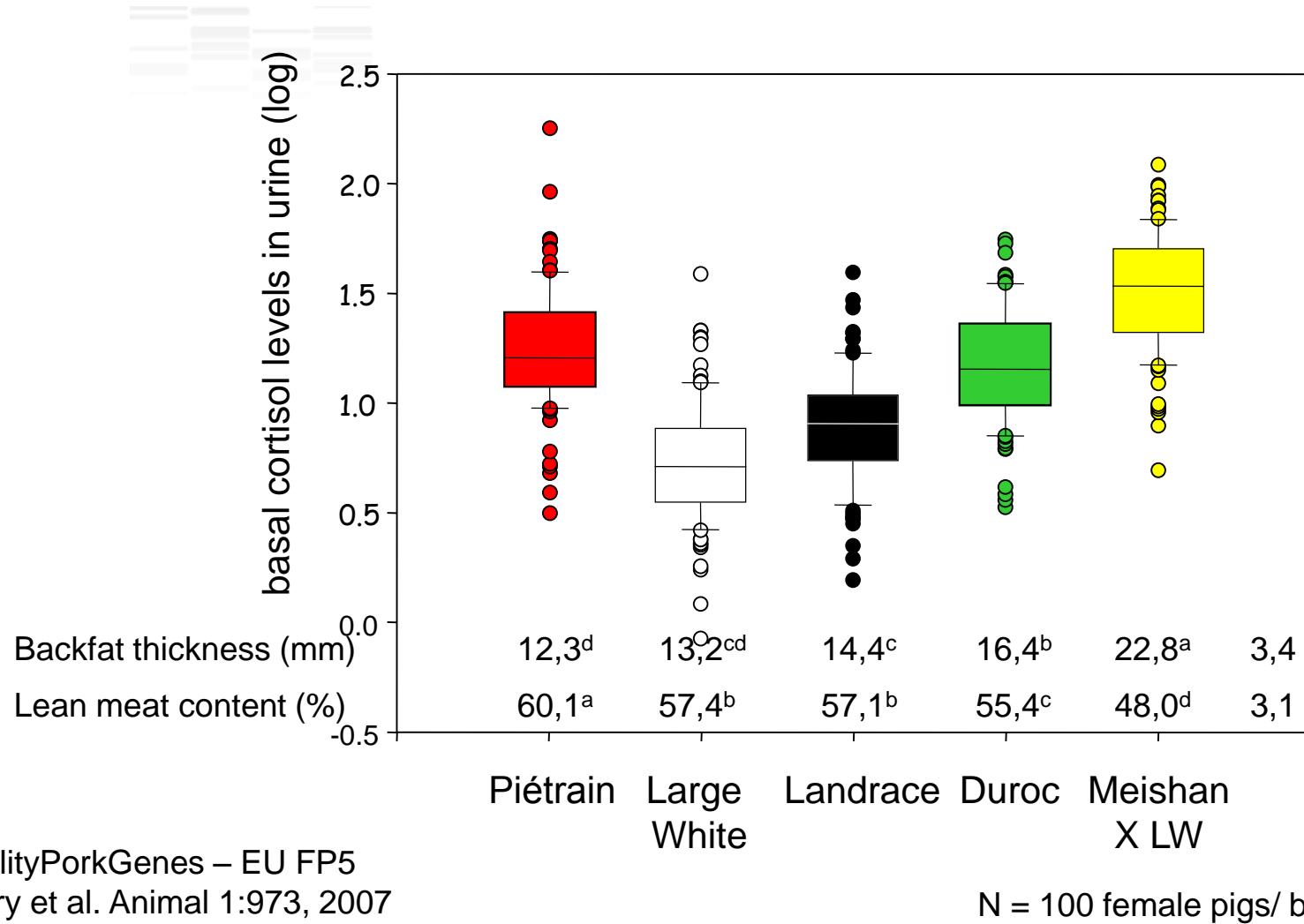
U. Weiler et al. / *Livestock Production Science* 54 (1998) 21–31



# Breeding objectives to increase robustness by selection of a stronger HPA axis

- ❖ Selection for fast growth rate, high feed efficiency and lean carcasses may have reduced HPA axis activity with negative consequences on robustness
- ❖ Working hypothesis : selection for a more active / reactive HPA axis could improve robustness with (hopefully) limited effects on production traits.
- ❖ Strong genetic influences on HPA axis activity
  - Differences between genetic stocks (e.g. Large White / Meishan)
  - Divergent genetic selection in chicken (ACTH, social stress), turkey (cold), duck (suspension), trout (confinement), mouse (restraint).
- ❖ Several sources of genetic variation
  - Cortisol production (sensitivity to ACTH)
  - Bioavailability
  - Receptor function

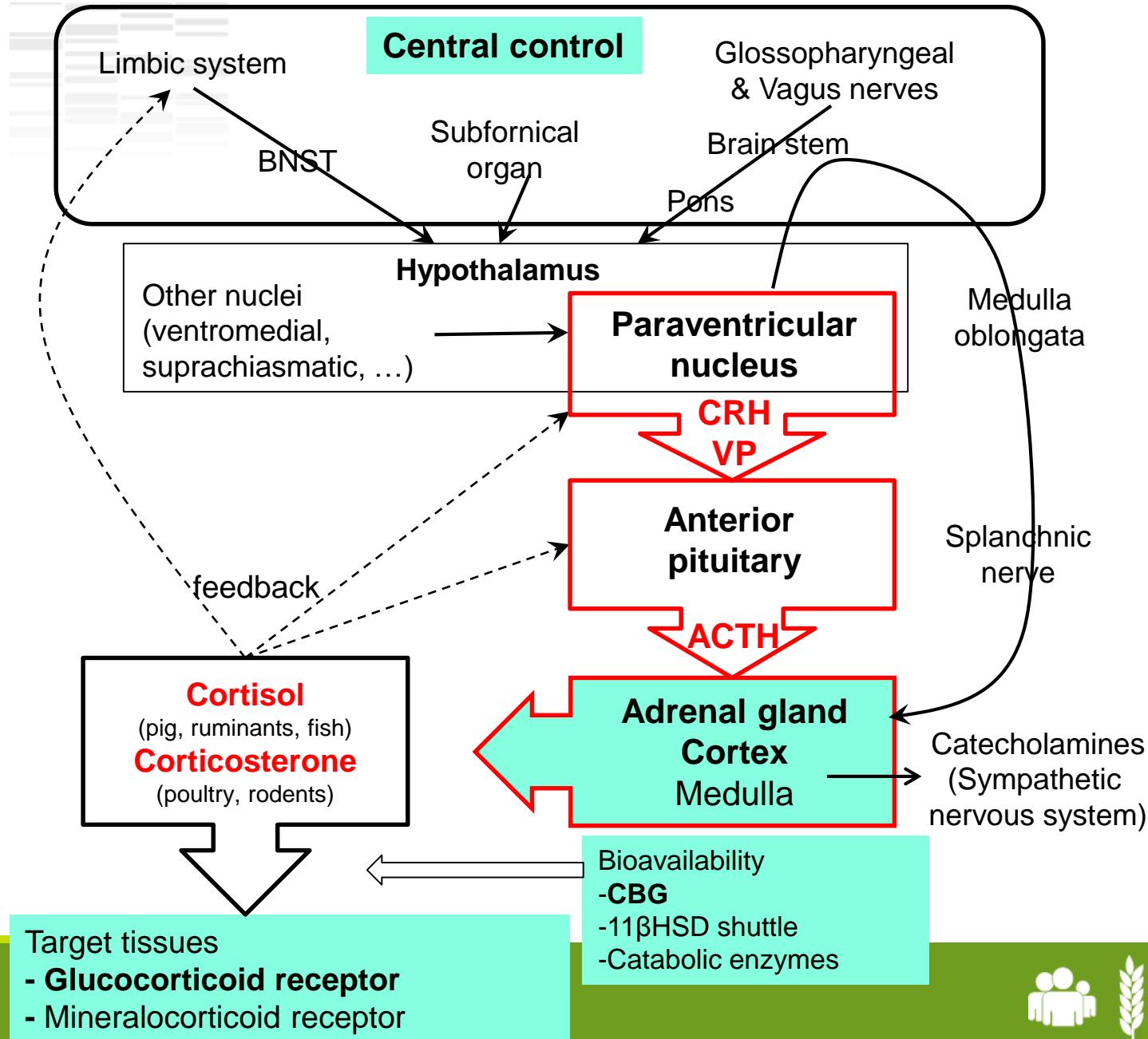
# Between- and within-breed variation of cortisol production



QualityPorkGenes – EU FP5  
Foury et al. Animal 1:973, 2007



# HPA axis, sources of genetic variability



# Sources of genetic variability in the HPA axis

## CORTICOSTEROID HORMONE PRODUCTION

- Response of the adrenal cortex to ACTH

- ❖ Is the main source of genetic variation in cortisol production
- ❖ Is an individual trait (stable across time)
  - Pigs    [Hennessy et al. 1988](#)
  - Humans - ‘Adrenal phenotype’    [Bertagna et al. 1994; Coste et al. 1994](#)
- ❖ Is heritable
  - $h^2 = 0.68$  - analysis in 30 litters of Large White pigs
    - [Larzul et al. Animal 9:1929, 2015](#)
  - Divergent selection
    - Direct selection in chicken, response to ACTH ([Edens et Siegel, 1975](#))
    - Main trait selected in response to stress (trout, duck)
- ❖ Molecular genomics
  - [Hazard et al. BMC Genomics 9:101, 2008](#)
  - [Jouffe et al. BMC Proceedings 3:S14, 2009](#)

# Adrenals: differentially expressed genes

## ❖ No difference

- In the **ACTH signaling pathway** (i.e. ACTH receptor, G protein, Adenylate Cyclase, Protein Kinase A and MAPK, ERK1, ERK2) except *Crem* (cAMP response element modulator) that activates genes involved in steroidogenesis
- In **steroidogenic enzymes** (i.e. *Cyp11a1*, *Cyp17*, *3 $\beta$ hsd*, *Cyp21*, *Cyp11b*)

## ❖ Differentially expressed

- **Cholesterol transport** (*Star*, *Ldlr*) and several kinases involved in phosphorylation of StAR (*Snf1lk*)
- **Tricarboxylic acid cycle** (*Mdh2*, *Sdha* and *Suclg2*) involved in heme biosynthesis (*Alas1* )
- Still to be characterized transcripts / genes

Hazard *et al.* *BMC Genomics* 9:101, 2008

# Sources of genetic variability in the HPA axis

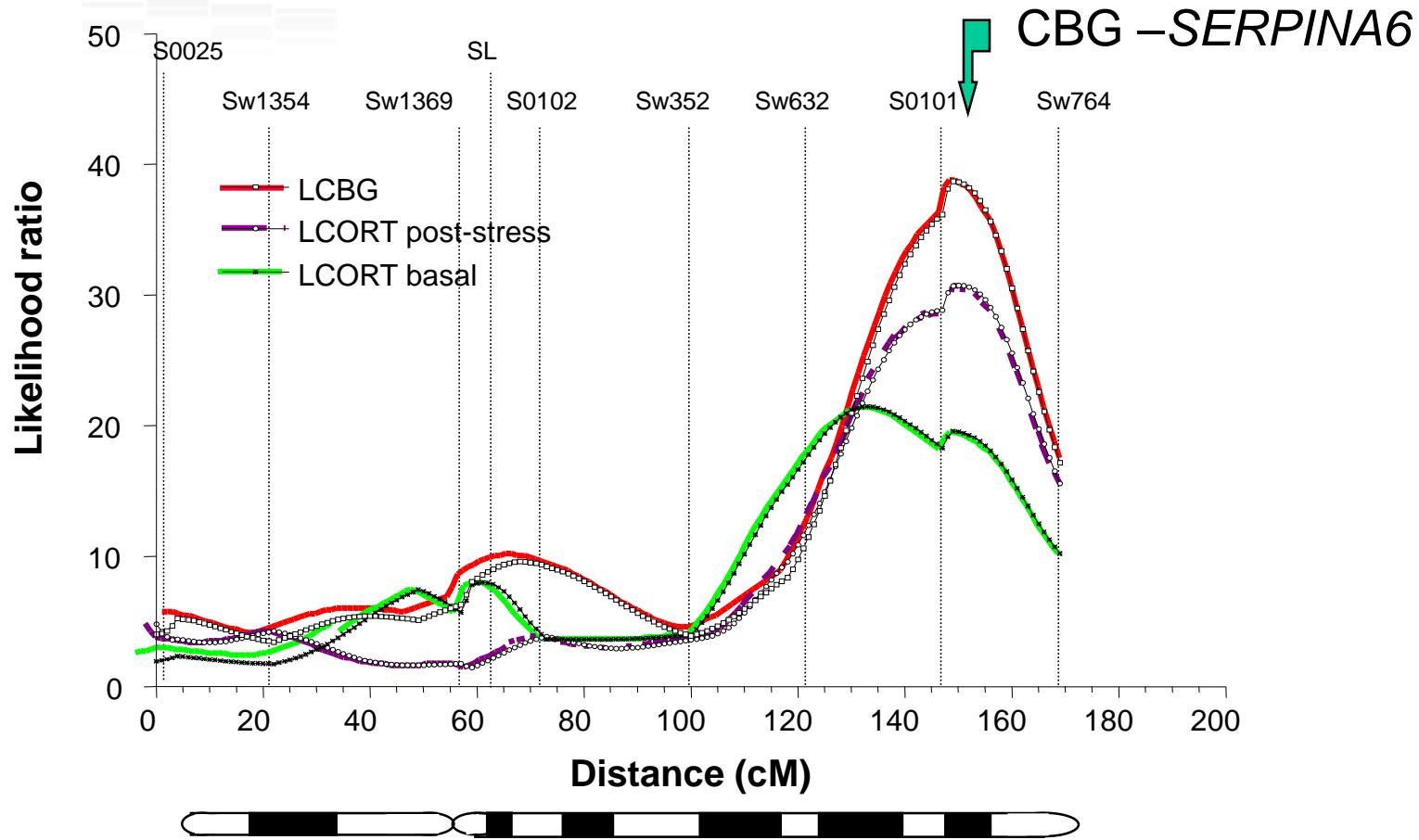
## BIOAVAILABILITY

- Corticosteroid-binding globulin (CBG)
- $11\beta$ -hydroxysteroid dehydrogenases

### ❖ Corticosteroid-binding globulin (CBG or transcortin)

- Glycoprotein synthesized in the liver and carrying cortisol in plasma  
Plays a dual role of sequestration of cortisol in plasma and delivery to the target tissues
  - Moisan Mol Cell Endocr 316:35, 2013
- Genetic determinant of cortisol levels by linkage study in a Large White x Meishan F2 population
  - Désautés et al. J Anim Sci 80:2276, 2002
  - Confirmed in rats (Solberg et al. 2006) and humans (Bolton et al. PLoS Genetics 2014)
- Molecular polymorphisms influencing protein properties **with consequences on adiposity and meat quality**
  - Ousova et al. Mol Endocrinol 18:1687, 2004
  - Geverink et al. J Anim Sci 84:204, 2006

# QTL cortisol / CBG on Ssr 7 (F2 LWxMS)



Désautés et al. J Anim Sci 80:2276 (2002)

Ousova et al. Mol Endo 18:1687 (2004)

# CBG and production traits

Pearson Correlation Coefficients between Carcass Composition Traits, Plasma CBG Capacity, and Total Plasma Control

	CBG	Cortisol (Basal)	Cortisol (Post Stress)
Backfat weight	r = 0.39 <i>P</i> = 0.014	r = -0.18 <i>P</i> = 0.28	r = 0.11 <i>P</i> = 0.50
% (Ham+ loin)	r = -0.40 <i>P</i> = 0.012	r = 0.18 <i>P</i> = 0.27	r = -0.11 <i>P</i> = 0.52
% (Back+ leaf fat)	r = 0.42 <i>P</i> = 0.008	r = -0.17 <i>P</i> = 0.29	r = 0.12 <i>P</i> = 0.45
Estimated muscle content	r = -0.43 <i>P</i> = 0.0064	r = 0.16 <i>P</i> = 0.33	r = -0.13 <i>P</i> = 0.41

Each cell of the table contains the Pearson correlation (*r*) coefficient and the probability (*P*). The number of observations is *n* = 39 in all cases.

# Sources of genetic variability in the HPA axis

## BIOAVAILABILITY

- Corticosteroid-binding globulin (CBG)
- $11\beta$ -hydroxysteroid dehydrogenases

- ❖ Corticosteroid-binding globulin (CBG or transcortin)
- ❖ Hormone metabolism
  - Cortisol – cortisone shuttle (hydroxysteroid dehydrogenases 1 & 2)
  - Local tissue bioavailability of cortisol may influence adiposity
  - Linkage/association between HSD11B1 polymorphism and carcass length and adiposity in pigs
    - Otieno et al. Anim Genet 36:36, 2004

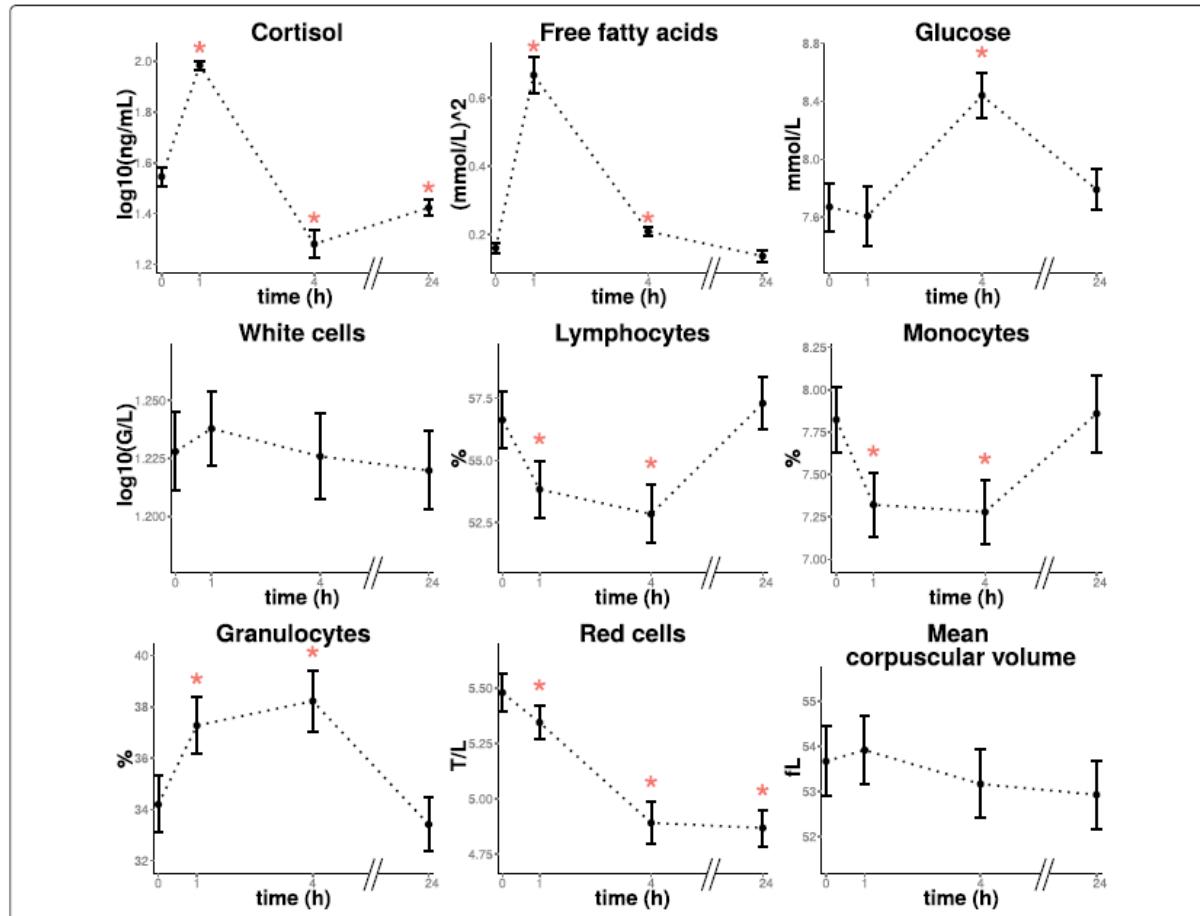
# Sources of genetic variability in the HPA axis

## ACTION ON TARGET TISSUES

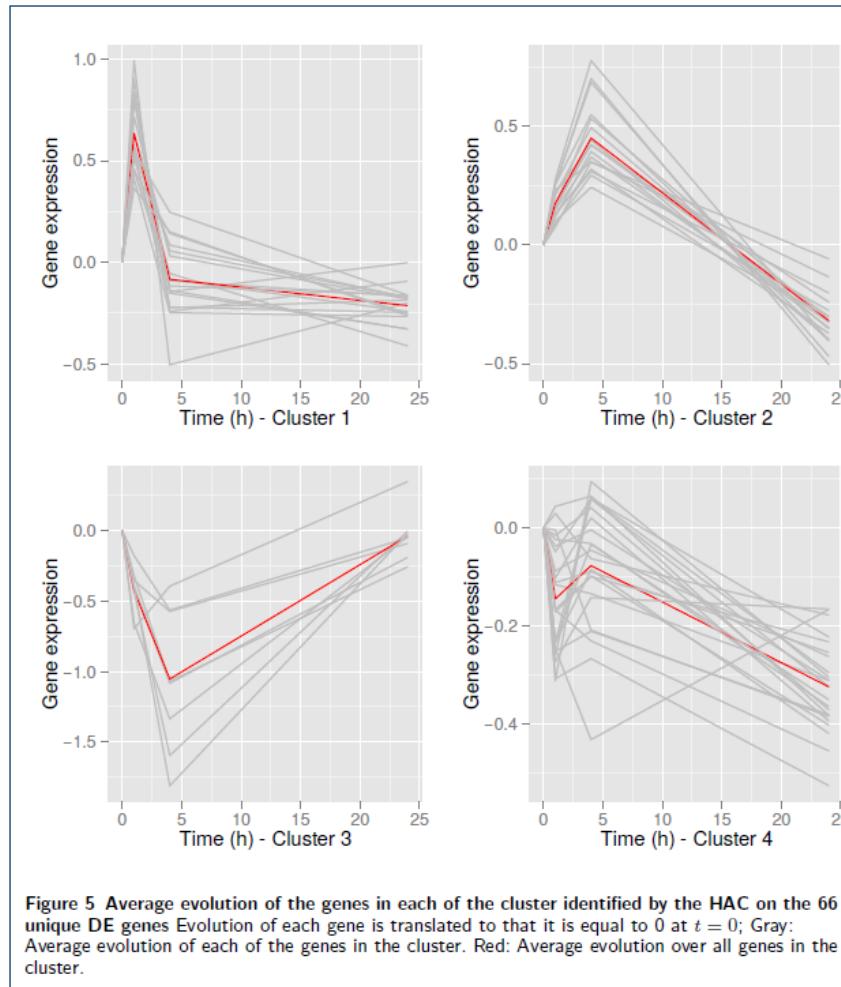
- Receptors or transduction pathways

- ❖ Cortisol receptors are transduction factors
  - Glucocorticoid receptors, GR, gene NR3C1
  - Mineralocorticoid receptors, MR, gene NR3C2
- ❖ Genetic influences and gene polymorphisms
  - well documented in humans and experimental animals
- ❖ Little information in farm animals
  - Functional differences LW/MS pigs  
Perreau et al. Life Sci 17:1501, 1999
  - Association studies (pigs)  
Murani et al. BMC Genetics 11:74, 2010; Terenina et al. DAE 44:81, 2013
  - Naturally occurring gain-of-function mutation Ala610Val  
Murani et al. 2012, 2016
- ❖ Functional genomic fingerprint of cortisol effects
  - (e.g. circulating white blood cells)

# Response to ACTH – biological measures in peripheral blood



# Response to ACTH - differentially expressed genes in circulating leucocytes



**Figure 5** Average evolution of the genes in each of the cluster identified by the HAC on the 66 unique DE genes Evolution of each gene is translated to that it is equal to 0 at  $t = 0$ ; Gray: Average evolution of each of the genes in the cluster. Red: Average evolution over all genes in the cluster.

# Phenotypic effect of the MAOA(-430A>T) SNP in the promoter region of monoamine oxidase A gene

## Association study

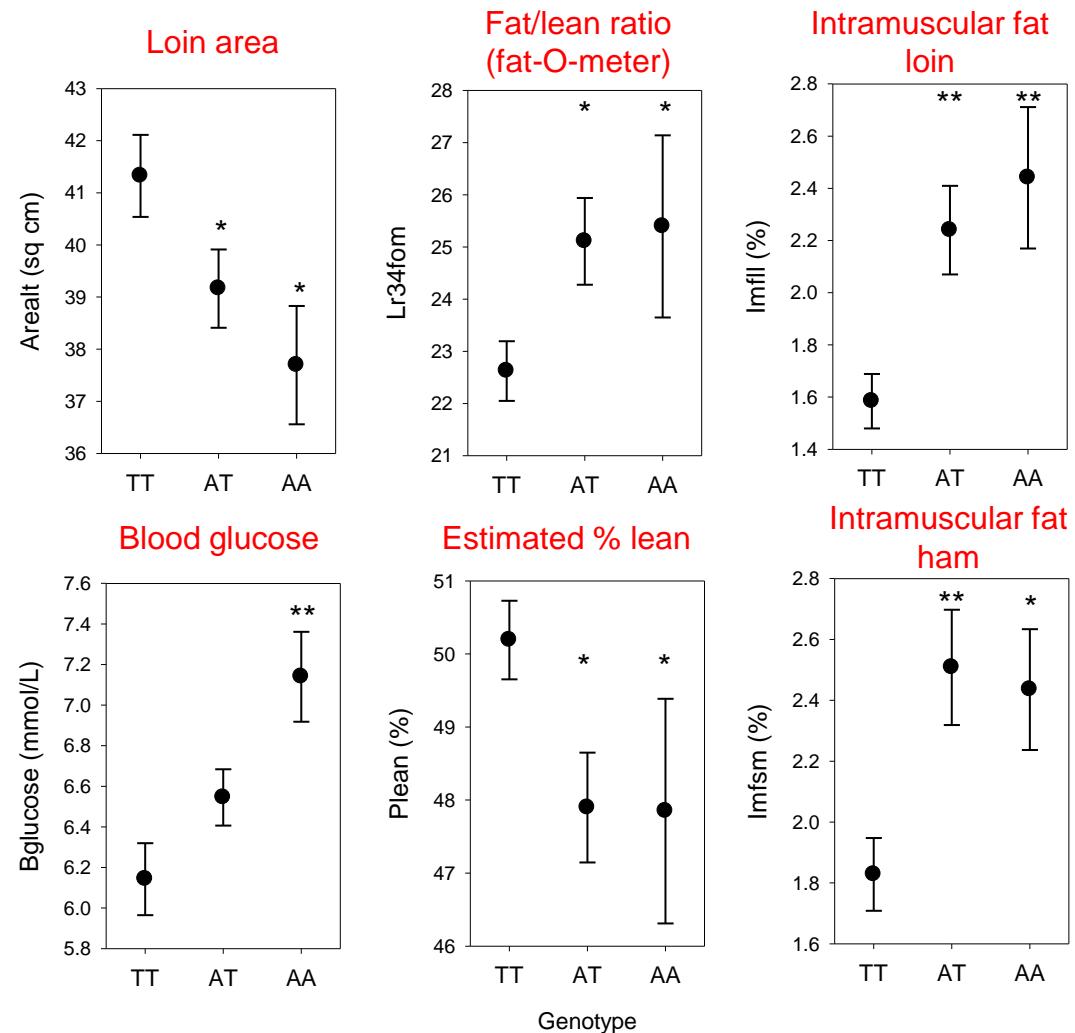
Female pigs ( $n = 100$ ) from an advanced intercross line (F9-F11) between Large White and Meishan

TT  $n = 40$

AT  $n = 41$

AA  $n = 11-13$

Terenina et al. DAE 44:81, 2013



# Phenotypic effect of the DRD3(-680C>T) SNP in the promoter region of dopamine receptor D3 gene

## Association study

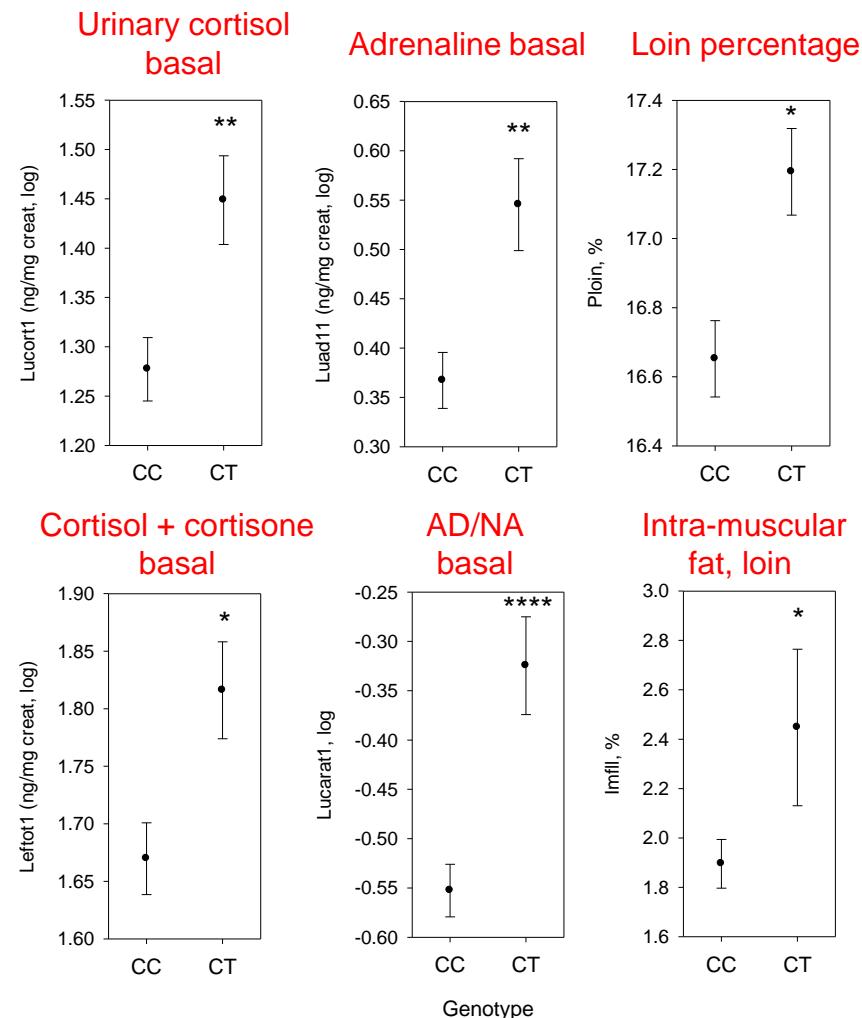
Female pigs ( $n = 100$ ) from an advanced intercross line (F9-F11) between Large White and Meishan

CC n = 66-68

CT n = 13-20

TT n = 3 (not shown)

Terenina et al. DAE 44:81, 2013



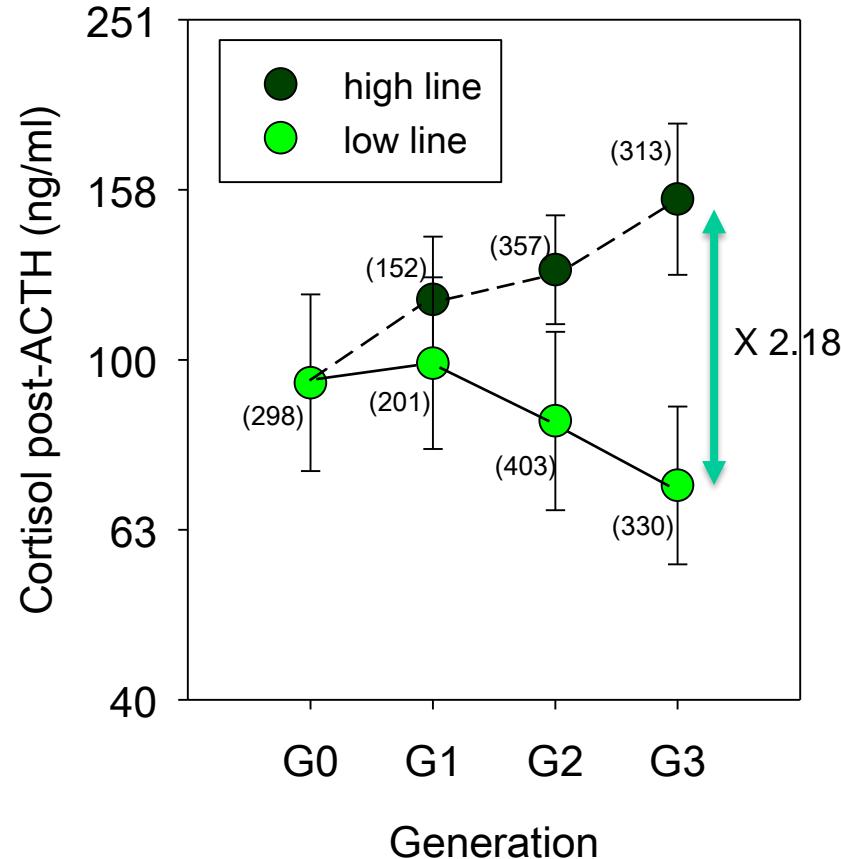
# HPA axis and robustness - proof of concept

## divergent selection for the response to ACTH in Large White pigs

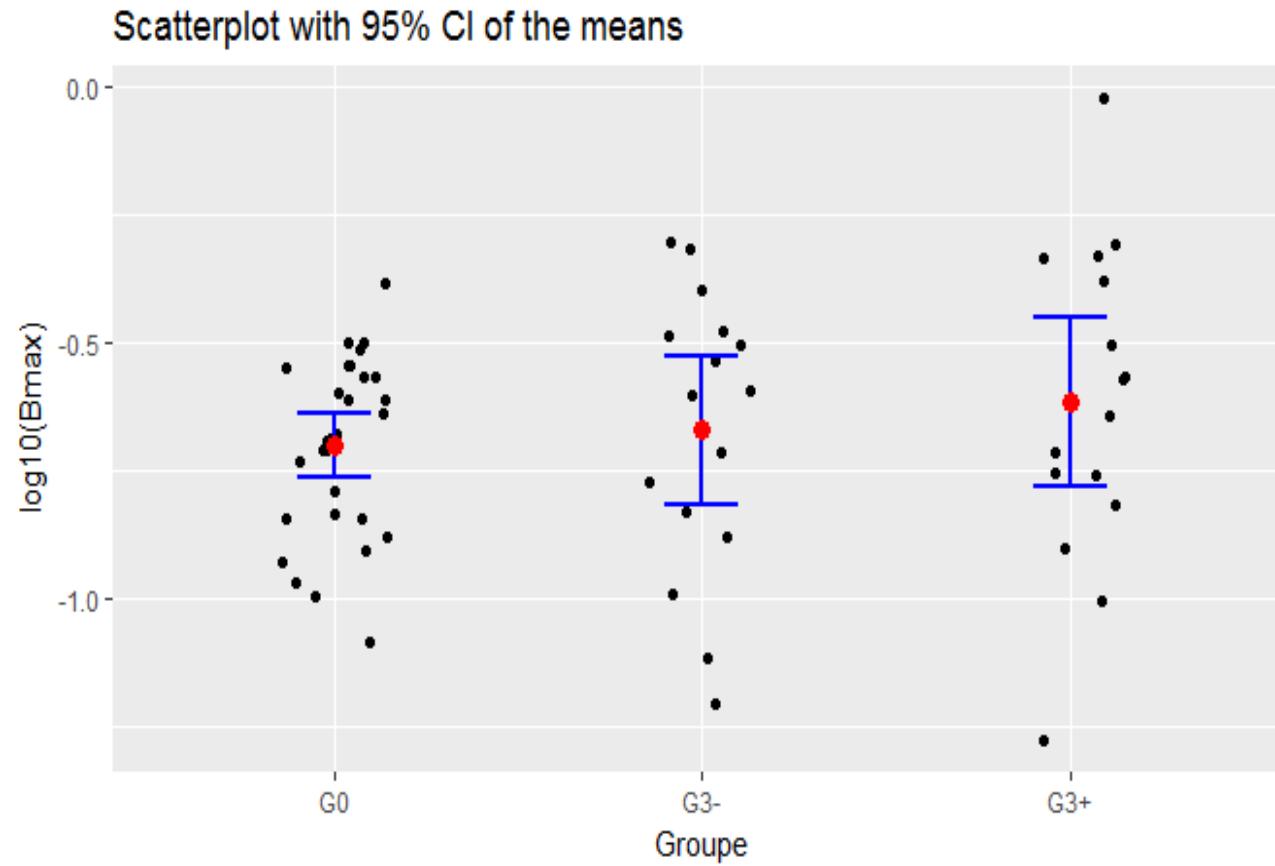


6 weeks

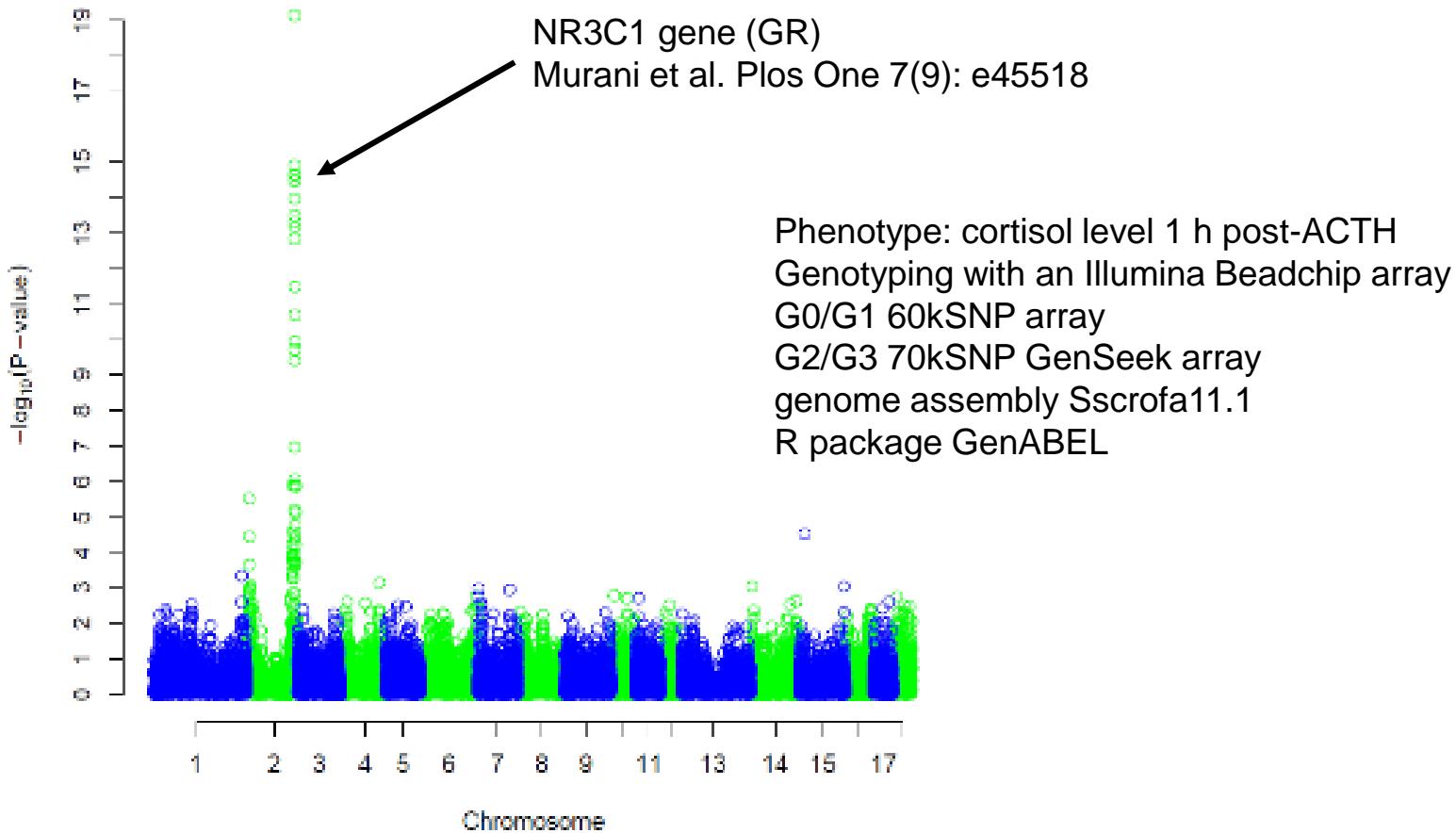
Realized heritability  $h^2 = 0.68 \pm 0.12$



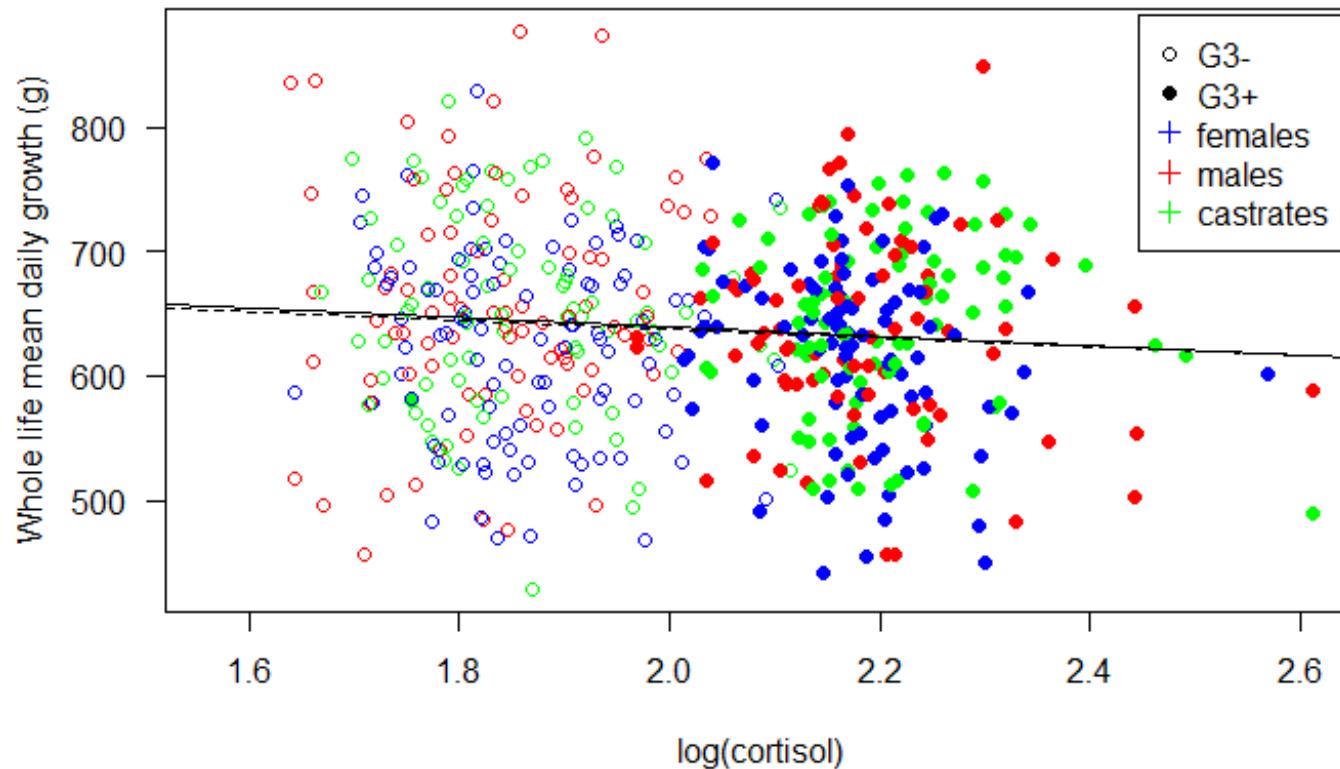
# CBG (cortisol-binding globulin) in selected lines



# Genome-wide association study



# Selected lines G3 – mean daily growth

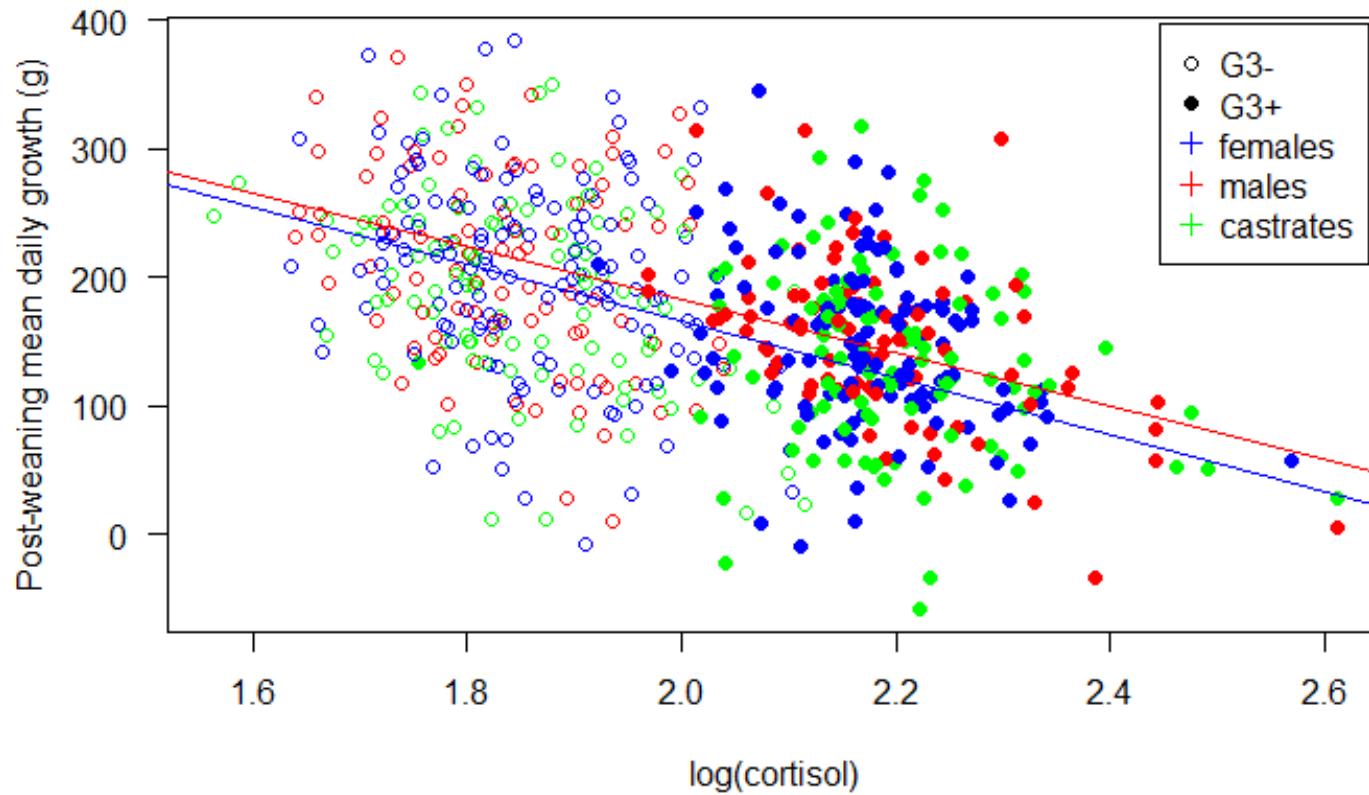


G3+ - G3- = -11 g/day, n.s.

Within groups, - 35 g/day per unit log(cortisol), n.s.

NO effect of selection  
NO influence of cortisol

# Selected lines G3 – post-weaning growth



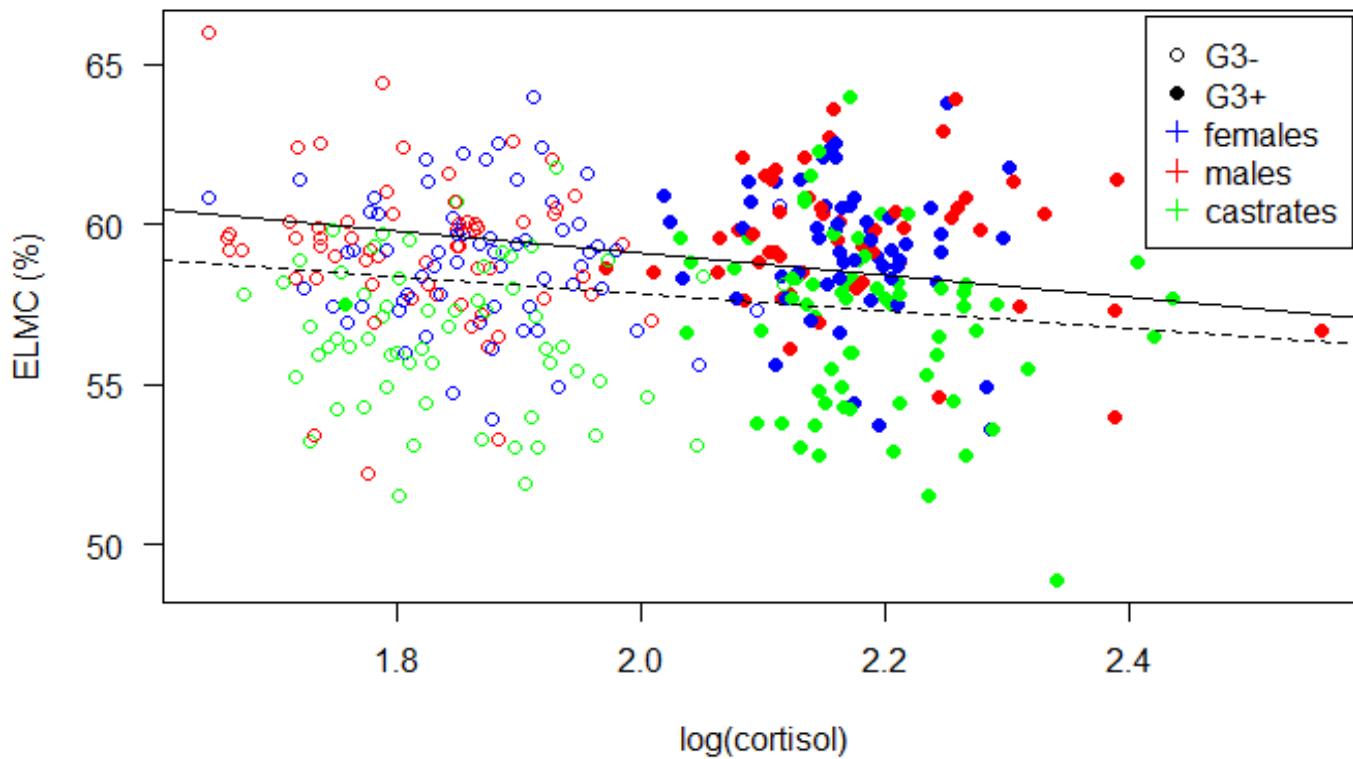
G3+ - G3- = -55 g,  $P < 2.10^{-16}$

Within groups, -214 g per unit log(cortisol),  $P < 2.10^{-15}$

At the same level of cortisol, G3+ - G3- = +16 g; n.s.

Strong effect of selection  
Related to cortisol levels

# Selected lines G3 – meat content



G3+ - G3- = 0.49%, P < 0.05

Within groups: ELMC -4.12% per unit log(cortisol), P < 0.002

At the same level of cortisol, G3+ - G3- = 1.90%, P < 0.0005

Small effect of selection  
Unrelated to cortisol levels

# Sources of genetic variability in the HPA axis

## CENTRAL DRIVE

- Inputs to the HPA axis
- CRH, vasopressin and receptors

## CORTICOSTEROID HORMONE PRODUCTION

- Response of the adrenal cortex to ACTH

## BIOAVAILABILITY

- Corticosteroid-binding globulin (CBG)
- $11\beta$ -hydroxysteroid dehydrogenases

## ACTION ON TARGET TISSUES

- Receptors or transduction pathways

## FUNCTIONAL CONSEQUENCES

Production / Meat quality / Robustness

**SYSTEMS  
GENETICS**

# FUNDING

INRA, Animal Genetics Division

ANR, Agence Nationale de la Recherche



- ❖ INRA, Genetics, Physiology and Breeding Systems, Toulouse
  - Elena Terenina, Catherine Larzul, Nathalie Villa-Vialaneix
  - Valérie Sautron (thesis)
  
- ❖ INRA, Genesi, Le Magneraud (experimental unit)
  - Yvon Billon

# Thank you for your attention



photo David McEnery

## Recent review papers

- Ann NY Acad Sci 1220:127, 2011
- Animal 5:651, 2011
- Dom Anim Endocr 43:116, 2012