# Androgenic response of *Triticum durum-Dasypyrum villosum* amphidiploids and their parental forms

H. Stoyanov 🖾, I. Belchev

Dobrudzha Agricultural Institute, General Toshevo, Bulgaria
Shpstoyanov@abv.bg

Abstract. Wide hybridization in cereal crops is one of the most efficient tools for the enrichment of genetic variability and addressing a number of breeding problems related to resistance and tolerance to biotic and abiotic stresses. Therefore, a large number of amphidiploids between species possessing different morphological, genetic and physiological properties have been developed. One of the most valuable species with regard to the possibilities for introducing valuable traits and properties into wheat species is the wild Dasypyrum villosum. With the aim to study the androgenic response of the Triticum durum-D. villosum amphidiploids, two accessions and their parental forms - the durum wheat cultivars Gergana and Argonavt and a landrace of the D. villosum - were studied. The following parameters were determined: callus induction, plant regeneration, yield of albino and green regenerants. It was found that the callus induction of the two studied amphidiploids differed significantly from that of the parental forms (2.1–7.2 %), being significantly higher, 30.7 and 16.5 %, respectively. Regardless of the difference in callus induction, the amphidiploids did not significantly differ from the parental forms in their regeneration ability. The yield of albino plants exceeded the yield of green regenerants and followed the tendency observed in callus induction. Green plants were found only in the amphidiploid Gergana-D. villosum and in the parental form durum wheat Gergana. Plants were regenerated from the species D. villosum, although they were only albinos, showing its good responsiveness to anther culture. The established characteristics of the amphidiploids and their parental forms make their practical use highly valuable for the improvement of different types of cereal crops. Key words: anther culture; and rogenic response; amphidiploid; Dasypyrum villosum; parental forms.

For citation: Stoyanov H., Belchev I. Androgenic response of *Triticum durum-Dasypyrum villosum* amphidiploids and their parental forms. *Vavilovskii Zhurnal Genetiki i Selektsii* = Vavilov Journal of Genetics and Breeding. 2022;26(2): 139-145. DOI 10.18699/VJGB-22-17

## Андрогенетическая реакция амфидиплоидов *Triticum durum-Dasypyrum villosum* и их родительских форм

Х. Стоянов 🖾, И. Белчев

Добруджанский сельскохозяйственный институт, г. Генерал Тошево, Болгария 🐵 hpstoyanov@abv.bg

> Аннотация. Отдаленная гибридизация злаков является одним из самых эффективных способов обогащения генетического разнообразия и решения множества селекционных задач в отношении устойчивости и толерантности к биотическому и абиотическому стрессу. Поэтому создано большое количество амфидиплоидов между отдельными видами, которые наделены разными морфологическими, генетическими и физиологическими характеристиками. Дикий вид Dasypyrum villosum – один из самых ценных видов с точки зрения возможности интродуцирования ценных качеств и свойств в разные сорта пшеницы. Для изучения реакции амфидиплоидов Triticum durum-D. villosum исследованы два образца, их родительские формы – сорта твердой пшеницы Гергана и Аргонавт, а также местная популяция вида D. villosum. Установлены следующие параметры: индукция каллусов, частота всех проростков к 100 эмбриоподобным структурам, частота альбиносных проростков и частота зеленых проростков. По полученным результатам выявлено, что индукция каллусов двух амфидиплоидов статистически достоверно отличается от родительских форм (2.1–7.2 %), будучи значительно выше, 30.7 и 16.5 % соответственно. Несмотря на разницу в индукции каллусов, амфидиплоиды практически не отличаются от родительских форм своей регенерирующей способностью. Частота альбиносных проростков значительно превышает частоту зеленых проростков, следуя тенденции, наблюдаемой в индукции каллусов. Зеленые проростки зарегистрированы только у амфидиплоида Гергана-D. villosum и у родительской формы твердой пшеницы Гергана. Растения вида D. villosum были регенерированы, и то, что были только альбиносные проростки, показывает хорошую отзывчивость вида на культивирование пыльников. Установленные характеристики испытанных амфидиплоидов и их родительских форм делают практическое использование этих амфидиплоидов особенно ценным для селекции различных видов злаков.

> Ключевые слова: пыльниковая культура; андрогенетическая реакция; амфидиплоид; *Dasypyrum villosum*; родительские формы.

### Introduction

The development of highly productive varieties of cultivated plants, which at the same time are characterized with stable yields and are resistant to different biotic and abiotic stress factors, is a primary task in plant breeding (Chahal, Gosal, 2000). However, the increase of yield within the genome of a given species is not limitless (Grassini et al., 2013). In this respect, there are different approaches to enrich the genome of the cultivated plants – wide hybridization, genetic engineering, genome editing technologies, etc. (Chahal, Gosal, 2000); Liu et al., 2014; Okada et al., 2019; Li, 2020; Wang et al., 2020).

Although contemporary science has reached high levels of use of the latter two technologies, wide hybridization remains a main conventional tool for achieving high genetic variability. There is a large amount of research on amphidiploids developed through wide hybridization among the cereal crops (Zhang et al., 2010; Ming et al., 2011; Babaiants et al., 2012; Stoyanov, 2013, 2014; Dai et al., 2015; Nemeth et al., 2015; Song et al., 2019; Klimushina et al., 2020; Zuo et al., 2020; Kiani et al., 2021). One of the most promising species for enrichment of the genome of common and durum wheat, however, is Dasypyrum villosum. This species has been described in detail with regard to the possibility of being used in the improvement work on the wheat species in the researches of A. Grądzielewska (2006a) and C. De Pace et al. (2011). In another research, A. Gradzielewska (2006b) described in detail a large number of studies on the production of hybrids, natural hybrids, substitution and addition lines with wheat and other species. There are a number of studies on the possibility of using the hybrids and amphidiploids of the wheat species with D. villosum in practical breeding (De Pace et al., 2001; Vaccino et al., 2010; De Pace et al., 2011; Zhang et al., 2015, 2016a, b, 2018; Ando et al., 2019). A. Stefani et al. (1987) reported rather detailed morphological characteristics of the amphidiploid Triticum durum-D. villosum.

Since plant breeding is a rather dynamic process, when developing lines from the cereal species, the biotechnological method of anther culture is often used to accelerate the breeding process (Belchev, 2003; Lantos, 2009). Different researchers report that the efficiency of the process and the production of a high number of green plants is related to the response to anther culture of the parental forms involved in the cross (Zamani et al., 2003; Dagüstü, 2008; Yildirim et al., 2008; El-Hennawy et al., 2011). In this respect, the developed amphidiploids, substitution and addition lines with D. villosum, are specific parental forms, the reaction to anther culture of which has not been studied up to now. The possibility to apply anther culture to amphidiploids in principle has been little investigated. The response to anther culture in the amphidiploids has been studied in the amphidiploid Aegilops variabilis-Secale cereale (Ponitka et al., 2002), and the authors determined 0.1-13.4 % of regenerants obtained from 100 androgenic embryoids. D. Plamenov et al. (2009) determined 1.9–3.2 % of green regenerants from 100 cultured anthers in the amphidiploid T. durum-T. monococcum ssp. aegilopoides. In tritordeum (Barcelo et al., 1994), it was also found out that anther culture is an efficient process. The results from these researches showed that different amphidiploids are able to give positive response to anther culture.

The aim of this study was to determine the reaction of the amphidiploid *T. durum-D. villosum* to anther cultivation in comparison to its parental forms.

#### Materials and methods

**Plant material.** Two accessions of the amphidiploid *T. durum-D. villosum* (1dv (Gergana-*D. villosum*) and 2dv (Argonavt-*D. villosum*)), a part of the collection of Dobrudzha Agricultural Institute were used, as well the durum wheat parental forms (*T. durum* cv. Gergana and cv. Argonavt) and the wild species *D. villosum*.

The accession of *D. villosum* (2n = 2x = 14 (VV)); family Poaceae, tribe Triticeae, subtribe Triticineae, genus *Dasypyrum*) was collected in Dobrich region in 2011.

Crosses Gergana  $\times D$ . *villosum* and Argonavt  $\times D$ . *villosum* were made conventionally, without embryo rescue in 2012; the obtained seeds (Gergana  $\times D$ . *villosum* – 3 seeds and Argonavt  $\times D$ . *villosum* – 8 seeds) were germinated and at tillering stage the plants (Gergana  $\times D$ . *villosum* – 1 plant and Argonavt  $\times D$ . *villosum* – 3 plants) were treated with colchicine in 2013. The seeds from the two obtained primary amphidiploids were multiplied several times.

Anther culture. The experiment was carried out during 2016/2017. Anther donor plants were grown under greenhouse conditions. The seeds from the accessions were germinated in Petri dishes and then planted in plastic pots. Fifteen plants from each accession were grown in three pots, using 10 plants per genotype. Primary, seedling were vernalized at 4 °C (3000 lx, 16 h day/8 h night) for 45 days. After this period, the plants were transferred to a cold greenhouse (5–15 °C) for about three months, and the temperature was later increased to 15–20 (25) °C. Tillers bearing spikes containing anthers with microspores at mid- to late uninucleate stage were cut, put in a vessel with water and pretreated at 4 °C for 8-9 days. Ten spikes from each genotype were collected. Cold pretreated spikes were surface sterilized with 70 % ethanol under aseptic conditions. Sixty anthers from each spike were placed in test tubes with 20 ml P2 induction medium (Chuang et al., 1978). The anthers were cultured at 28 °C in darkness for about 60 days. After the 30th day, they were periodically checked for induction of embryogenic structures (calli and embryoids), which were transferred to test tubes with 10 ml regeneration medium (Zhuang, Jia, 1983) and cultured at 25 °C (3000 lx, 16 h day/8 h night). Green and albino regenerants were counted after 30 days.

The androgenic response was estimated by the following traits: callus induction (CI) (number of embryogenic structures induced per 100 cultured anthers, %), plant regeneration (PR) (number of regenerated green and albino plants per 100 embryogenic structures, %), frequency (yield) of green plants (YGR) (number of regenerated green plants per 100 cultured anthers, %) and frequency (yield) of albino plants (YAR) (number of regenerated albino plants per 100 cultured anthers, %).

**Statistics.** The obtained results were summarized over genotypes and parameters. One way ANOVA was carried out with the aim of determining the effect of the genotype on the studied parameters to estimate their androgenic response. Significant differences between the amphidiploids and their

parental forms were calculated based on the Duncan test. To process the data, software MS Office Excel 2003 was used, and to perform ANOVA and the Duncan test – IBM SPSS Statistics v.19.

#### Results

The results on the androgenic potential of the investigated amphidiploids and their parental forms (Table 1) showed that accession 1dv had the highest callus induction (30.7 %), and durum wheat Argonavt – the lowest (2.1 %). Between the parental forms, there were no significant differences (both between the durum wheat cultivars and between the species durum wheat and *D. villosum*). The two amphidiploids differed significantly by their callus induction, which was probably related to the effect of the maternal component.

The yield of green plants, averaged for the entire investigated set, was extremely low. In the entire experiment, only 5 green regenerants were produced, one of them being from the durum wheat cultivar Gergana, and the other 4 - from the amphidiploid 1dv (Gergana-*D. villosum*). No green regenerants were obtained from cultivar Argonavt and from the amphidiploid 2dv. Also, no green plants were produced from the wild species *D. villosum*. Although there was a rather small number of the obtained plants for formulating a general tendency for the effect of the parental forms, the presence of green plants in cultivar Gergana and the amphidiploid, in which it was involved, was probably due to genotypic specificity.

The albino plants considerably exceeded the green regenerants. In practice, they were predominant with regard to the total number of regenerants. The amphidiploid 1dv again had the highest yield of albino plants (10.3 %), and the lowest values were observed in cultivar Argonavt (0.4%). The tendency in yield of obtained albinos largely followed the tendency of callus induction. The two amphidiploids significantly differed from the parental forms by their values, as well as between themselves (10.3 and 5.8 %, respectively). Meanwhile, significant differences between the two durum wheat cultivars and between the durum wheat and the wild species were not registered. The higher yield values of the albino plants in the amphidiploid 1 dv may be related to the higher responsiveness of cultivar Gergana, which was the maternal component of this amphidiploid, although the difference between Gergana and Argonavt was not significant.

On the whole, plant regeneration, expressed as a number of regenerants per 100 embryogenic structures, was comparatively low. The highest values were read in the two investigated aphidiploids (35.9 and 35.4 %, respectively), and the lowest in the wild species D. villosum (13.0 %). This parameter did not follow the tendency observed in the values of callus induction and yield of green and albino regenerants. There were no significant differences between any of the studied accessions. However, higher plant regeneration was registered in the amphidiploids, in comparison to cultivar Argonavt and the wild species, and the difference with cultivar Gergana was considerably lower. The differences not being significant was an indication that the regeneration potential of all studied accessions was practically identical, and the differences formed were entirely random. The total number of regenerants, however, expressly followed the tendency of callus induction and yield of albino plants. The higher responsiveness to anther culture of the two investigated amphidiploids in comparison to either of the parental forms could be clearly observed in this parameter.

The results from the analysis of the variance of the studied parameters (Table 2) showed that the genotype had a significant effect on the parameters callus induction and yield of albino regenerants. This allows supposing that the separate accessions gave specific responses and that there are significant differences between them, as determined by the Duncan test that was carried out. At the same time, the effect of the separate accessions on the plant regeneration and the yield of green plants was not significant. Worth mentioning are accessions Gergana and Gergana-*D. villosum*, in which higher responsiveness to anther culture was observed, in general. Nevertheless, these results do not give a definite answer to the question of whether the amphidiploids are different as a biologically distinct organism from the two parental forms with regard to their androgenic response.

When summarizing the results at the level of the species, a clear tendency of the amphidiploid *T. durum-D. villosum* having significantly higher callus induction and yield of albino regenerants was evident (Table 3).

Simultaneously, significant differences between the two parental forms were not observed, the values of both parameters being significantly lower in them. The yield of green plants from the parental forms and from the amphidiploid was extremely low and did not allow forming a clear tendency. In this case, the production of green regenerants was rather random, without observing significant differences between the investigated species. Plant regeneration, at the levels of

Table 1. Androgenic response of parental forms and durum wheat-D. villosum amphidiploids
--

ianie in indiogenie i									
Genotype	NCA	NOC	CI, %	NRC	PR, %	NAR	YAR, %	NGR	YGR, %
Gergana	600	43	7.2ab	13	30.2a	12	2ab	1	0.2ab
Argonavt	480	10	2.1a	2	20.0a	2	0.4a	0	0.0a
D. villosum (Dv)	540	23	4.3ab	3	13.0a	3	0.6a	0	0.0a
1dv (Gergana-Dv)	600	184	30.7c	66	35.9a	62	10.3c	4	0.7b
2dv (Argonavt-Dv)	480	79	16.5b	28	35.4a	28	5.8b	0	0.0a

Note. NCA – number of cultivated anthers; NOC – number of obtained calli; CI – callus induction; NRC – number of regenerative calli; PR – plant regeneration; NAR – number of albino regenerants; YAR – yield of albino regenerants; NGR – number of green regenerants; YGR – yield of green regenerants.

Parameters		Sum of squares	df	Mean square	F	Significance	
Cl	Between groups	0.51892	4	0.12973	8.693	0.000	
	Within groups	0.59697	40	0.01492			
	Total	1.11589	44				
YAR	Between groups	0.06774	4	0.01694	7.799	0.000	
	Within groups	0.08686	40	0.00217			
	Total	0.15460	44				
YGR	Between groups	0.00032	4	0.0008	2.159	0.091	
	Within groups	0.00147	40	0.00004			
	Total	0.00179	44				
PR	Between groups	0.36055	4	0.09014	0.899	0.474	
	Within groups	4.01134	40	0.10028			
	Total	4.37188	44				

Table 2. ANOVA according to factor "accession" of the studied accessions

Note. CI - callus induction; YAR - yield of albino regenerants; YGR - yield of green regenerants; PR - plant regeneration.

Table 3. Androgenic response of parental forms and durum wheat-D. villosum amphidiploid

Species	NCA	NOC	CI, %	NRC	PR, %	NAR	YAR, %	NGR	YGR, %
<i>T. durum</i> (Td)	1080	53	4.9a	15	28.3a	14	1.3a	1	0.1a
D. villosum (Dv)	540	23	4.3a	3	13.0a	3	0.6a	0	0.0a
Td-Dv	1080	263	24.4b	94	35.7a	90	8.3b	4	0.4a

Note. NCA – number of cultivated anthers; NOC – number of obtained calli; CI – callus induction; NRC – number of regenerative calli; PR – plant regeneration; NAR – number of albino regenerants; YAR – yield of albino regenerants; NGR – number of green regenerants; YGR – yield of green regenerants.

Parameters		Sum of squares	df	Mean square	F	Significance	
CI	Between groups	0.41771	2	0.20885	12.564	0.000	
	Within groups	0.69818	42	0.01662			
	Total	1.11589	44				
YAR	Between groups	0.05763	2	0.02881	12.480	0.000	
	Within groups	0.09698	42	0.00231			
	Total	0.15460	44				
YGR	Between groups	0.00011	2	0.00005	1.349	0.271	
	Within groups	0.00168	42	0.00004			
	Total	0.00179	44				
PR	Between groups	0.31393	2	0.15696	1.625	0.209	
	Within groups	4.05795	42	0.09662			
	Total	4.37188	44				

Note. CI – callus induction; YAR – yield of albino regenerants; YGR – yield of green regenerants; PR – plant regeneration.

both genotype and species, did not differ as a tendency. The observed differences were not significant (see Tables 3 and 4), which indicated that the studied amphidiploid did not differ from the parental forms by its regeneration capacity.

#### Discussion

Concerning the results obtained on the androgenic response of the used accessions, it should be emphasized, that no source was found in world literature that would present data on the amphidiploid *T. durum-D. villosum* or the species *D. villosum*. An exception was the research of X. Chen et al. (1996), who suggested applying the anther culture method on hybrids (not amphidiploids) of the  $F_1$  (*T. durum* × *D. villosum*). These authors reported successful production of amphidiploids, regenerated from colchicine-treated calli. At the same time, there are researches on the use of tissue cultures on three-

component hybrids *T. aestivum*  $\times$  (*T. durum-D. villosum*). H. Li et al. (2005) reported lines with high powdery mildew resistance obtained from such hybrids through the method of embryo rescue and subsequent anther culture.

D. Plamenov et al. (2009), when investigating the androgenic response of accessions from the amphidiploid T. durum-T. monococcum ssp. aegilopoides, came up with results different from ours. The reported callus induction was 3.3-11.7 % for the two studied accessions, the plant regeneration was considerably higher, 33.8-68.4 %, respectively, and the albino regenerants yield was 1.9-3.2 %. At the same time, the yield of green plants (0.4–0.8 %) was a little higher than the data we obtained in our experiment (0.0-0.7 %). These authors reported a total of seven regenerated plants from both accessions, this parameter being significant, unlike the results we obtained. Using anther culture in the amphidiploid Ae. variabilis-S. cereale, and P2 medium, A. Ponitka et al. (2002) observed 1.4-15.7 % of callus induction, and on C17 medium - 20.0-65.2 %. Subsequently, the authors reported 0.1-13.4 % yield of green regenerants using 190-2 regeneration medium. It was found out that the androgenic response was strongly dependent on the genotype, similar to the results of the experiment we conducted. Successful regeneration of green plants through the method of anther culture has also been reported for an aneupolyhaploid of Thynopyrum ponticum (Wang et al., 1991), for the amphidiploid Festuca pratensis-Lolium multiflorum (Lesniewska et al., 2001; Zwierzykowski et al., 2001; Rapacz et al., 2005) and the amphidiploid Cyclamen persicum-C. purpurascens (Ishizaka, 1998).

In contrast to these results, the parental forms were characterized with much lower androgenic response. This was confirmed by the absence of callus induction in *Ae. variabilis* and rye, reported by A. Ponitka et al. (2002), and also in the species *T. monococcum* ssp. *aegilopoides* in the research of D. Plamenov et al. (2009). Durum wheat is also characterized by weak androgenic response, in general. M. Doğramacı-Altuntepe et al. (2001), using 10 durum wheat genotypes, obtained only 248 green regenerants from 86,400 anthers (0.29%). F. J'Aiti et al. (1999), investigating 15 durum wheat genotypes and 7500 cultivated anthers, obtained just three albino regenerants and one green plant.

L. Cistúe et al. (2006), on the other hand, reported significantly higher production of green plants, but including 6-benzylaminopurine or 6-furfurilaminopurine in the induction medium (C17). In more recent researches, the production of haploids, even by the method of isolated microspores, has been of extremely low efficiency in durum wheat (Slama-Ayed et al., 2019). These results entirely corresponded to the data we obtained with regard to the two cultivars Argonavt and Gergana. Clear genotypic specificity was observed in the better response of Gergana to anther culture as compared to Argonavt, although the difference was not statistically significant. It is probable that this tendency is the reason for the amphidiploid Gergana-D. villosum having better responsiveness to anther culture. In this respect, the amphidiploid T. durum-D. villosum we investigated, and the amphidiploids reported by A. Ponitka et al. (2002) and D. Plamenov et al. (2009) were closer by their androgenic response to the response of triticale (which is a typical amphidiploid crop) than to the response of the parental forms. J. Pauk et al. (2000), K. Marciniak et al.

(2003), C. Lantos et al. (2014) and H. Stoyanov et al. (2019) demonstrated that in triticale the albino regenerants are often predominant, similar to the amphidiploid we studied. The values of the green regenerants in triticale also varied (from 0.9 to 27.9 %, but more often within 3–6 %), according to data from various researches (Gonzales, Jouve, 2000; Marciniak et al., 2003; Banaszak, 2011; Lantos et al., 2014).

In contrast to the above responses of the parental forms Ae. variabilis, S. cereale and T. monococcum ssp. aegilopoides, our study, although limited in volume, demonstrated the comparatively good responsiveness of the species D. villosum to anther culture. This is the first time when results on regenerants from this species (although only albinos) are being reported. At the same time, it should be emphasized that until this moment results from testing of the reaction of D. villosum to the anther culture method have never been reported. This is highly significant for the breeding of the wheat species since it would allow transferring genes from the wild species through the methods of wide hybridization and anther culture more easily, quickly and efficiently. X. Chen et al. (1996) and C. Li et al. (2000) reported common wheat lines resistant to powdery mildew, which were obtained by crossing common wheat to the amphidiploid T. durum-D. villosum, followed by embryo rescue and anther culture. Such results showed that the combination of wide hybridization with the method of anther culture is an efficient tool that can be used in the breeding of different cereal crops.

#### Conclusion

Based on the presented results, the following conclusions could be made:

- 1. For the first time, results on the androgenic response (callus induction, plant regeneration, yield of albino plants, yield of green plants) of the amphidiploid *T. durum-D. villosum* and of the parental component *D. villosum* are being reported.
- 2. The callus induction of the two studied amphidiploids differed significantly from that of the parental forms (2.1-7.2 %), being considerably higher 30.7 and 16.5 %, respectively.
- 3. The plant regeneration of the investigated accessions varied within a certain range (13.0–35.9 %), the differences not being statistically significant. This indicated that in spite of the differences in the callus induction, the amphidiploids did not practically differ from the parental forms by their regeneration capacity.
- 4. Although plant regeneration was observed in all studied accessions, the yield of albino plants considerably exceeded the yield of green regenerants and followed the tendency observed in callus induction the two amphidiploids had significantly higher values. At the same time, green plants were registered only in the amphidiploid Gergana-*D. villosum* and in the parental form durum wheat Gergana. Such results emphasized the genotypic specificity of the response to anther culture.
- 5. Plants were regenerated from the species *D. villosum*, although only albinos, which indicated its good responsiveness to anther culture. This, together with the good response of the amphidiploids with the participation of this species, makes their practical use, in combination with the anther culture method, highly valuable for improving the cereals.

#### References

- Ando K., Krishnan V., Rynearson S., Rouse M.N., Danilova T., Friebe B., See D., Pumphrey M.O. Introgression of a novel Ug99effective stem rust resistance gene into wheat and development of *Dasypyrum villosum* chromosome-specific markers via genotypingby-sequencing (GBS). *Plant Dis.* 2019;103:1068-1074. DOI 10.109/ PI-S-05-18-0831-RE.
- Babaiants O.V., Babaiants L.T., Horash A.F., Vasil'ev A.A., Trackovetskaia V.A., Paliasn'iii V.A. Genetics determination of wheat resistance to *Puccinia graminis* f. sp. *tritici* deriving from *Aegilops cylindrica*, *Triticum erebuni* and amphidiploid 4. *Tsitol. Genet.* 2012; 46(1):10-17. (in Ukrainian)
- Banaszak Z. Breeding of triticale in DANKO. In: Tagungsband der 61. Jahrestagung der Vereinigung der Pflanzenzüchter und Saatgutkaufl eute Österreichs 23–25 November 2010. Raumberg-Gumpenstein. Irdning, 2011;65-68.
- Barceló P., Cabrera A., Hagel C., Lörz H. Production of doubled-haploid plants from tritordeum anther culture. *Theor: Appl. Genet.* 1994; 87:741-745.
- Belchev I. Studies on Anther Culture of Common Winter Wheat (*Triticum aestivum* L.) and Application of Double Haploid Lines in Breeding. General Toshevo, 2003. (in Bulgarian)
- Chahal G.S., Gosal S.S. Principles and Procedures of Plant Breeding: Biotechnological and Conventional Approaches. New York: CRC Press, 2000.
- Chen X., Xu H.J., Du L.P., Shang L.M., Han B., Shi A., Xiao S. Transfer of gene resistant to powdery mildew from *H. villosa* to common wheat by tissue culture. *Sci. Agric. Sin.* 1996;29:1-8.
- Chuang C.C., Ouyang T.W., Chia H., Chou S.M., Ching C.K. A set of potato media for wheat anther culture. In: Proc. of Symposium on Plant Tissue Culture 1978. Peking: Science Press, 1978;51-56.
- Cistué L., Soriano M., Castillo A.M., Vallés M.P., Sanz J.M., Echávarri B. Production of doubled haploids in durum wheat (*Triticum turgidum* L.) through isolated microspore culture. *Plant Cell Rep.* 2006;25(4):257-264. DOI 10.1007/s00299-005-0047-8.
- Dagüstü N. Diallel analysis of anther culture response in wheat (*Triticum aestivum* L.). *Afr. J. Adv. Biotechnol.* 2008;7(19):3419-3423.
- Dai S., Li Z., Xue X., Jia Y., Liu D., Pu Z., Zheng Y., Yan Z. Analysis of high-molecular-weight glutenin subunits in five amphidiploids and their parental diploid species *Aegilops umbellulata* and *Aegilops uniaristata*. *Plant Genet. Resour.* 2015;13(2):186-189. DOI 10.1017/S1479262114000719.
- De Pace C., Snidaro D., Ciaffi M., Vittori D., Ciofo A., Cenci A., Tanzarella O.A., Qualset C.O., Mugnozza G.T.S. Introgression of *Dasypyrum villosum* chromatin into common wheat improves grain protein quality. *Euphytica*. 2001;117(1):67-75. DOI 10.1023 A:100 4095705460.
- De Pace C., Vaccino P., Cionini P.G., Pasquini M., Bizzarri M., Qualset C.O. Dasypyrum. Chapter 4. In: Kole C. (Ed.). Wild Crop Relatives: Genomic and Breeding Resources. Cereals. Berlin; Heidelberg: Springer-Verlag, 2011;185-292. DOI 10.1007/978-3-642-14228-4 4.
- Doğramacı-Altuntepe M., Peterson T.S., Jauha P.P. Anther culture-derived regenerants of durum wheat and their cytological characterization. J. Hered. 2001;92(1):56-64.
- El-Hennawy M.A., Abdalla A.F., Shafey S.A., Al-Ashkar I.M. Production of doubled haploid wheat lines (*Triticum aestivum* L.) using anther culture technique. *Ann. Agric. Sci.* 2012;56(2):63-72. DOI 10.1016/j.aoas.2011.05.008.
- Gonzales J.M., Jouve N. Improvement of anther culture media for haploid production in triticale. *Cereal Res. Commun.* 2000;28(1-2): 65-72.
- Grądzielewska A. The genus *Dasypyrum* part 2. *Dasypyrum villosum* a wild species used in wheat improvement. *Euphytica*. 2006a; 152:441-454.
- Grądzielewska A. The genus *Dasypyrum* part 1. The taxonomy and relationships within *Dasypyrum* and with *Triticeae* species. *Euphytica*. 2006b;152:429-440.

- Grassini P., Eskridge K., Cassman K. Distinguishing between yield advances and yield plateaus in historical crop production trends. *Nat. Commun.* 2013;4:2918. DOI 10.1038/ncomms3918.
- Ishizaka H. Production of microspore-derived plants by anther culture of an interspecific F1 hybrid between *Cyclamen persicum* and *C. purpurascens. Plant Cell Tissue Organ Cult.* 1998;54:21-28.
- J'Aiti F., Benlhabib O., Sharma H.C., El Jaafari S., El Hadrami I. Genotypic variation in anther culture and effect of ovary coculture in durum wheat. *Plant Cell Tissue Organ Cult.* 1999;59:71-76.
- Kiani R., Arzani A., Meibody S.A.M.M., Rahimmalek M., Razavi K. Morpho-physiological and gene expression responses of wheat by *Aegilops cylindrica* amphidiploids to salt stress. *bioRxiv*. 2021. DOI 10.1101/2020.06.07.139220.
- Klimushina M.V., Kroupin P.Y., Bazhenov M.S., Karlov G.I., Divashuk M.G. Waxy gene-orthologs in wheat × *Thinopyrum* amphidiploids. *Agronomy*. 2020;10(7):963. DOI 10.3390/agronomy100 70963.
- Lantos C. *In vitro* androgenezis induction in wheat (*Triticum aesti-vum* L.), Triticale (×*Triticosecale* Wittmack), spice pepper (*Capsi-cum annuum* L.) and integration of the results into breeding: Thesis of the Ph.D. dissertation. Szent Istvan University, 2009.
- Lantos C., Bona L., Boda K., Pauk J. Comparative analysis of *in vitro* anther- and isolated microspore culture in hexaploid Triticale (×*Tri-ticosecale* Wittmack) for androgenic parameters. *Euphytica*. 2014; 197(1):27-37.
- Lesniewska A., Ponitka A., Slusarkiewicz-Jarzina A., Zwierzykowska E., Zwierzykowski Z., James A.R., Thomas H., Humphreys M.W. Androgenesis from *Festuca pratensis* × *Lolium multiflorum* amphidiploid cultivars in order to select and stabilize rare gene combinations for grass breeding. *Heredity*. 2001;86:167-176.
- Li C. Breeding crops by design for future agriculture. J. Zhejiang Univ. Sci. B. 2020;21(6):423-425. DOI 10.1631/jzus.B2010001.
- Li H., Chen X., Xin Z.Y., Ma Y.Z., Xu H.J., Chen X.Y., Jia X. Development and identification of wheat–*Haynaldia villosa* T6DL.6VS chromosome translocation lines conferring resistance to powdery mildew. *Plant Breed*. 2005;124:203-205. DOI 10.1111/j.1439-0523. 2004.01062.x.
- Li H.J., Li Y.W., Zhang Y.M., Li H., Guo B.H., Wang Z.N., Wen Z.Y., Liu Z.Y., Zhu Z.Q., Jia X. Tissue culture induced translocation conferring powdery mildew resistance between wheat and *Dasypyrum villosum* and its marker-assisted selection. *Yi Chuan Xue Bao*. 2000;27(7):608-613.
- Liu D., Zhang H., Zhang L., Yuan Z., Hao M., Zheng Y. Distant hybridization: a tool for interspecific manipulation of chromosomes. In: Pratap A., Kumar J. (Eds.). Alien Gene Transfer in Crop Plants. Vol. 1. New York: Springer, 2014. DOI 10.1007/978-1-4614-8585-8\_2.
- Marciniak K., Kaczmarek Z., Adamski T., Surma M. The anther-culture response of triticale line × tester progenies. *Cell. Mol. Biol. Lett.* 2003;8:343-351.
- Ming D., Xiuqin W., Fu Q., Zhao Q., Zhao F., Wang Y. Cytogenetical analysis of hybrid F1 from common wheat and *Aegilops ventricosa×Aegilops cyiindrica* amphidiploid. *Agric. Sci. Technol. Hunan.* 2011;12(9):1298-1302.
- Nemeth C., Yang C.-y., Kasprzak P., Hubbart S., Scholefield D., Mehra S., Skipper E., King I., King J. Generation of amphidiploids from hybrids of wheat and related species from the genera *Aegilops*, *Secale*, *Thinopyrum*, and *Triticum* as a source of genetic variation for wheat improvement. *Genome*. 2015;58(2):71-79. DOI 10.1139/gen-2015-0002.
- Okada A., Arndell T., Borisjuk N., Sharma N., Watson-Haigh N.S., Tucker E.J., Baumann U., Langridge P., Whitford R. CRISPR/Cas9mediated knockout of Ms1 enables the rapid generation of malesterile hexaploid wheat lines for use in hybrid seed production. *Plant Biotechnol. J.* 2019;17:1905-1913. DOI 10.1111/pbi.13106.
- Pauk J., Poulimatka M., Toth K.L., Monostori T. *In vitro* androgenesis of triticale in isolated microspore culture. *Plant Cell Tissue Organ Cult.* 2000;61(3):221-229. DOI 10.1023/A:1006416116366.

- Plamenov D., Belchev I., Spetsov P. Anther culture response of *Triticum durum* × *T. monococcum* ssp. *aegilopoides* amphiploid. *Cereal Res. Commun.* 2009;37(2):255-259. DOI 10.1556/CRC.37.2009.2.13.
- Ponitka A., Slusarkiewicz-Jarzina A., Wojciechowska B. Production of haploids and doubled haploids of the amphiploids *Aegilops variabilis × Secale cereale. Cereal Res. Commun.* 2002;30(1):39-45. DOI 10.1007/BF03543387.
- Rapacz M., Gąsior D., Humphreys M.W., Zwierzykowski Z., Płażek A., Lésniewska-Bocianowska A. Variation for winter hardiness generated by androgenesis from *Festuca pratensis × Lolium multiflorum* amphidiploid cultivars with different winter susceptibility. *Euphytica*. 2005;142:65-73.
- Slama-Ayed O., Bouhaouel I., Ayed S., De Buyser J., Picard E., Amara H.S. Efficiency of three haplomethods in durum wheat (*Triticum turgidum* subsp. *durum* Desf.): isolated microspore culture, gynogenesis and wheat × maize crosses. *Czech J. Genet. Plant Breed*. 2019;55(3):101-109. DOI 10.17221/188/2017-CJGPB.
- Song Z., Dai S., Jia Y., Zhao L., Kang L., Liu D., Wei Y., Zheng Y., Yan Z. Development and characterization of *Triticum turgidum– Aegilops umbellulata* amphidiploids. *Plant Genet. Resour.* 2019; 17(1):24-32. DOI 10.1017/S1479262118000254.
- Stefani A., Meletti P., Onnis A. Morphological characteristics of the experimental allopolyploid *Triticum durum* × *Haynaldia villosa* (2n = 42). Can. J. Bot. 1987;65:1948-1951.
- Stoyanov H. Status of remote hybrids in the Poaceae: problems and prospects. Agric. Sci. Technol. 2013;5(1):3-12.
- Stoyanov H. Analysis and assessment of amphidiploids of Triticum-Aegilops group as a source of genetic diversity. *Bulg. J. Agric. Sci.* 2014;20(Suppl. 1):173-178.
- Stoyanov H., Belchev I., Baychev V. Changes in the androgenic response of triticale (*×Triticosecale* Wittm.) as a result of the inclusion of sweet potato extract in the induction media. *Rastenievadni Nauki*. 2019;56(5):92-98. (in Bulgarian)
- Vaccino P., Banfi R., Corbellini M., De Pace C. Improving the wheat genetic diversity for end-use grain quality by chromatin introgression from the wheat wild relative *Dasypyrum villosum*. Crop Sci. 2010;50(2):528-540.
- Wang H., Sun S., Ge W., Zhao L., Hou B., Wang K., Lyu Z., Chen L., Xu S., Guo J., Li M., Su P., Li X., Wang G., Bo C., Fang X., Zhuang W., Cheng X., Wu J., Dong L., Chen W., Li W., Xiao G., Zhao J., Hao Y., Xu Y., Gao Y., Liu W., Liu Y., Yin H., Li J., Li X., Zhao Y., Wang X., Ni F., Ma X., Li A., Xu S.S., Bai G., Nevo E., Gao C., Ohm H., Kong L. Horizontal gene transfer of *Fhb7* from fungus underlies *Fusarium* head blight resistance in wheat. *Science*. 2020; 368(6493):eaba5435. DOI 10.1126/science.aba5435.
- Wang R.R.C., Marburger J.E., Hu C.J. Tissue-culture-facilitated production of aneupolyhaploid *Thinopyrum ponticum* and amphidi-

ploid *Hordeum violaceum* × *H. bogdanii* and their uses in phylogenetic studies. *Theor. Appl. Genet.* 1991;81:151-156. DOI 10.1007/BF00215716.

- Yildirim M., Bahar B., Genç İ., Hatipoğlu R., Altıntaş S. Reciprocal effects in anther cultures of wheat hybrids. *Biol. Plant.* 2008;52:779-782. DOI 10.1007/s10535-008-0152-y.
- Zamani I., Gouli-Vavdinoudi E., Kovacs G., Xynias I., Roupakias D., Barnabas B. Effect of parental genotypes and colchicine treatment on the androgenic response of wheat F1 hybrids. *Plant Breed*. 2008; 122(4):314-317. DOI 10.1046/j.1439-0523.2003.00866.x.
- Zhang L.Q., Liu D.C., Zheng Y.L., Yan Z.H., Dai S.F., Li Y.F., Jiang Q., Ye Y.Q., Yen Y. Frequent occurrence of unreduced gametes in *Triticum turgidum–Aegilops tauschii* hybrids. *Euphytica*. 2010;172:285-294. DOI 10.1007/s10681-009-0081-7.
- Zhang R., Fan Y., Kong L., Wang Z., Wu J., Xing L., Cao A., Feng Y. Pm62, an adult-plant powdery mildew resistance gene introgressed from *Dasypyrum villosum* chromosome arm 2VL into wheat. *Theor. Appl. Genet.* 2018;131(12):2613-2620. DOI 10.1007/s00122-018-3176-5.
- Zhang R., Feng Y., Li H., Yuan H., Dai J., Cao A., Xing L., Li H. Cereal cyst nematode resistance gene *CreV* effective against *Heterodera filipjevi* transferred from chromosome 6VL of *Dasypyrum villosum* to bread wheat. *Mol. Breed.* 2016a;36:122. DOI 10.1007/s11032-016-0549-9.
- Zhang R., Sun B., Chen J., Cao A., Xing L., Feng Y., Lan C., Chen P. Pm55, a developmental-stage and tissue-specific powdery mildew resistance gene introgressed from *Dasypyrum villosum* into common wheat. *Theor. Appl. Genet.* 2016b;129:1975-1984. DOI 10.1007/ s00122-016-2753-8.
- Zhang R.Q., Hou F., Feng Y.G., Zhang W., Zhang M.Y., Chen P.D. Characterization of *a Triticum aestivum–Dasypyrum villosum* T2VS·2DL translocation line expressing a longer spike and more kernels traits. *Theor. Appl. Genet.* 2015;128(12):2415-2425. DOI 10.1007/s00122-015-2596-8.
- Zhuang J.J., Jia X. Increasing differentiation frequencies in wheat pollen callus. In: Cell and Tissue Culture Techniques for Cereal Crop Improvement. Beijing: Science Press, 1983.
- Zuo Y., Xiang Q., Dai S., Song Z., Bao T., Hao M., Zhang L., Liu G., Li J., Liu D., Wei Y., Zheng Y., Yan Z. Development and characterization of *Triticum turgidum–Aegilops comosa* and *T. turgidum– Ae. markgrafii* amphidiploids. *Genome*. 2020;63(5):263-273. DOI 10.1139/gen-2019-0215.
- Zwierzykowski Z., Ponitka A., Slusarkiewicz-Jarzina A., Zwierzykowska E., Lesniewska-Bocianowska A. Androgenesis in amphidiploid F1 hybrids and cultivars of Festulolium. *Zeszyty Problemowe Postepow Nauk Rolniczych (Poland)*. 2001;474:47-54. (in Polish)

#### ORCID ID

H. Stoyanov orcid.org/0000-0003-0733-9077

Conflict of interest. The authors declare no conflict of interest.

Received August 2, 2021. Revised November 27, 2021. Accepted December 30, 2021.